From: Jay Field

Eric Blischke/R10/USEPA/US@EPA To:

Robert Gensemer; Burt Shephard/R10/USEPA/US@EPA; Chip Humphrey/R10/USEPA/US@EPA; Joe Cc:

Goulet/R10/USEPA/US@EPA; Robert Neely

Re: Summary of Sediment Bioassay Interpretation Resolution Subject:

07/14/2009 01:35 PM Date:

Eric.

my understanding was that unlike the round 2 data, the round 3 represented retesting of the same sediment so that there were results for each of the 2 batches of R3 tox tests. If the R3 "replicate" samples were actually 100 ft apart, then not averaging is appropriate as Bob states. note that the R2 6 stations with samples collected 100 ft apart were labelled differently and several of those field replicates are included as separate samples in the ref envelope. Jay

Robert Gensemer wrote:

Eric: I don't think anyone is suggesting that we average the upstream samples with the three samples per location. I don't exactly know what is different about G788, but I assume the a/b samples were splits from the same physical sample? I would average the latter, but not the former. -Bob

----Original Message----

From: Blischke.Eric@epamail.epa.gov [mailto:Blischke.Eric@epamail.epa.gov] Sent: Tuesday, July 14, 2009 12:26 PM

To: Jay Field

Cc: Shephard.Burt@epamail.epa.gov; Humphrey.Chip@epamail.epa.gov; Goulet.Joe@epamail.epa.gov; Robert Gensemer; Robert Neely

Subject: Re: Summary of Sediment Bioassay Interpretation

Resolution

When we designed the original upstream sampling program, we identified

six locations and collected three samples at each location. However,

the samples were collected some distance (~100') apart as I recall.

Given this fact, would we really be introducing bias by including the

duplicate? Would it really matter if we collected three or four samples

from a given location? Your argument assumes that the duplicates have

the same characteristics beyond the similar characteristics of each of

the six locations or the upstream area in general.

Eric

Jay Field <Jay.Field@noaa.

aov>

То

Blischke/R10/USEPA/US@EPA 07/14/2009 12:17

CC

DMRobert Gensemer

Eric

<rgensemer@parametrix.com>,

Burt

Shephard/R10/USEPA/US@EPA,

Chip

Humphrey/R10/USEPA/US@EPA,

Joe

Goulet/R10/USEPA/US@EPA,

Robert

Neely

<Robert.Neely@noaa.gov>

Subject

Bioassay

irrelevant if

Re: Summary of Sediment
Interpretation Resolution

Eric,
Regarding number 4): It does matter, since it gives more weight
to two
of the stations in the reference envelope. You could include both
but
give them a weight of 1/2, if your curve fitting package allows
weighting. Helping the curve fitting procedure is kind of

the distribution is skewed by including two samples twice. Jay $% \left(1\right) =\left(1\right) +\left(1\right) +\left($

Blischke.Eric@epamail.epa.gov wrote:

Bob, thanks for the quick response. I have a few questions/comments:

Regarding number 2), do we understand why the biomass values don't match. If the control normalization was done correctly and there were no reporting errors, could there be a difference in how total biomass

was

reported?
Regarding number 3), I will make sure that I specify survivorship in

my

email.
Regarding number 4), it seems we did not specify whether to pool or to handle to duplicates as individual sample results when calculating the reference envelope. My question is two-fold - 1) does it matter? and

2)

if we include the duplicates as individual samples, could this help

curve fitting procedure because we now have an additional one or two samples?
Regarding number 6) Burt and I discussed this. He seemed to think

that

it is more valid statistically to fit the entire curve rather than the lower end due to the small number of samples at the lower end of the distribution. My original thought was along the lines of yours but

Burt

convinced me otherwise. We can revisit this though. Once I get some additional feedback, I will finalize the email and

send

to John Toll and Bob Wyatt.
Thanks, Eric

Robert Gensemer

<rgensemer@param

etrix.com>

То

Eric

Blischke/R10/USEPA/US@EPA,

07/13/2009 08:34 Shephard/R10/USEPA/US@EPA, Burt

PM "jay.field@noaa.gov"

<jay.field@noaa.gov>, Joe

Goulet/R10/USEPA/US@EPA

CC

Chip Humphrey/R10/USEPA/US@EPA

Subject

RE: Summary of Sediment Bioassay

Interpretation

Resolution

Eric: A few observations from my perspective:

2) The control-normalization looks correct for biomass, but if $\ensuremath{\mathrm{I}}$

recall

(I don't have my files with me at the moment) that LWG's

biomass

values

for individual stations did not quite match values that Jay derived

for

table RE-1.
3) You have the control normalization correct
(test/control) but we

need

to be careful to recommend use of survivorship, not mortality, to be fully consistent with our guidance and numeric examples. I realize

Table

2-1 used mortality, but we have been very consistent all along that we need to use survivorship, and from a recent call with Burt, Don McD. agrees that control-normalized survivorship is the correct value to

use,

not ctrl-norm mortality. Yes, they relate directly (or should I say, inversely) to one another, but the 5th percentile calculation could be different using one vs. the other, so we need to be consistent, and

use

survivorship.
4) I could not find any explicit guidance regarding the duplicate RE samples. Its not in the McDonald report that I can find, and I don't think we went into this level of detail in the problem formulation. It may be one of those things that just seemed very obvious to all of us, and so never felt the need to explicitely direct it. Actually, it may have only come up, to my recollection, during our own RE calculations

in

March. So table RE-1 definitely reflects this approach,

don't

think it was spelled out in the text. 6) I agree with your summary here, except to say that we need to not just chose the best overall curve fit, but particularly in the case of Hyalella biomass, we need a curve that fits the lower tail (i.e., 5th %ile) of the distribution best. For the other three endpoints, this is probably not an issue (i.e., best fit is also best 5th %ile fit). But for Hyl biomass, we need to think more carefully about what

distribution

fits at the lower tail of the distribution. I think this is a valid approach that makes the best out of the available data. LWG's curve

fit

created a 5th %ile value that was quite a bit lower than the empirical numbers; I do not think that was the most appropriate representation

 $\circ f$

the data.

Bob

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p Before printing, please think green.

From: Blischke.Eric@epamail.epa.gov
[Blischke.Eric@epamail.epa.gov]
Sent: Monday, July 13, 2009 5:02 PM
To: Robert Gensemer; Shephard.Burt@epamail.epa.gov;

jay.field@noaa.gov;

Goulet.Joe@epamail.epa.gov

rgensemer@parametrix.com

Cc: <u>Humphrey.Chip@epamail.epa.gov</u> Subject: Summary of Sediment Bioassay Interpretation Resolution

As you are aware, we have been discussing some of the details of the LWG's interpretation of the Portland Harbor sediment bioassay results. Some elements of the interpretation were discussed during a conference call on Thursday, June 18, 2009.

Here is where I believe we are:

1) No transcription errors were identified during a review of the reference envelope bioassay results. The total biomass calculations were done correctly. 2) 3) Mortality should be computed as test/control. This is consistent with Table 2-1 in the March 17, 2006 Bioassay Interpretation Report, ASTM Method E-1706, and EPA Guidance. 4) Duplicate reference envelope samples should be pooled (averaged) rather than treated as individual samples. This is consistent with February 15, 2008 problem formulation (Note: is this the correct

reference? I could not find this in either the problem

nor

formulation

the MacDonald benthic risk evaluation)
5) Identification of Level 1, Level 2 and Level 3 thresholds: The toxicity thresholds should be calculated based on 10% of the reference envelope not an absolute 10%. This is consistent with Tables RE 1,

RE-2

Calculation and
Use of Reference Envelope for Portland Harbor Sediment
Toxicity Test
Interpretation
6) Identification of the 5% of the reference envelope
should be
accomplished using a range of curve fitting procedures
appropriate for
the data set distribution. The curve fitting procedure
with the best
overall fit should be selected and the 5% calculated
using the best

and the text of EPA's March 31, 2009 direction on the

fit

curve fitting procedure.

The above procedures for computing the results of the bioassay tests, calculating hit/no-hit designations, developing the reference envelope

and identifying Level 1, Level 2 and Level 3 toxicity hits should be followed.

Please look this over and make sure it matches up with the recommended procedures. See also my note about the pooling of the reference duplicate samples. Once everyone agrees with the outlined procedures,

I

will send an email to the LWG summarizing this and recommending a conference call to discuss if there area any questions.

Thanks, Eric

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